

PHLOROTANNINE AUS *HIMANTHALIA ELONGATA**

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Key Word Index—*Himantalia elongata*; brown algae; Fucaceae; polyhydroxyphenols.

In der Fraktion der acetylierten Phenole aus *Himantalia elongata* (L.) S. F. Gray ist bisher Phloroglucin-triacetat (1) dc nachgewiesen worden [1].

1 kg gefriergetrocknete Alge (Roscoff, Bretagne 1973) wurde in gewohnter Weise mit EtOH extrahiert, die Phenole mit EtOAc ausgeschüttelt (Ausbeute 700 mg, 0,07%) und mit Ac₂O-Py acetyliert [2]. Im DC der acetylierten Phenole (Kieselgel F₂₅₄, CHCl₃-Me₂CO (9:1)) findet sich außer 1 eine Reihe weiterer UV-Licht löschender mit Vanillin-H₂SO₄ anfärbarer Zonen: 2 (*R_f* 0,81), 3 (*R_f* 0,74), 4 (*R_f* 0,59), 5 (*R_f* 0,51) und andere mehr. 2 und 3 wurden über eine Kieselgelsäule mit CHCl₃ gereinigt und aus MeOH umkristallisiert.

2 (4 mg, Smp 113–114°) ergibt im MS ein Molekülion bei *m/e* 460, von dem bis zu fünfmal Bruchstücke mit 42 Masseneinheiten bis zu einem Ion bei *m/e* 250 abspalten. Weitere charakteristische Fragmentationen finden sich bei *m/e* 43 (MeCO⁺), 69 (C₃HO₂⁺), 126 (C₆H₆O₃⁺) und 142 (C₆H₆O₄⁺). Aufgrund der spektralen Daten, des Smp und des DC-Vergleichs in verschiedenen Fließmitteln ist 2 mit dem zuerst in *Cystoseira tamariscifolia* [3] nachgewiesenen Diphlaretholpentaacetat (2,4,6,3',5'-Pentaacetoxydiphenyläther) identisch.

Für 3 (6 mg, Smp 177,5–178,5°) wurde das Molekülion bei *m/e* 502 gefunden. Die Verbindung spaltet bis zu sechsmal Fragmente mit 42 Masseneinheiten bis zu einem Ion bei *m/e* 250 ab und ist aufgrund der spektralen Daten, des Smp und des chromatographischen Verhaltens identisch mit dem zuvor aus *Fucus vesiculosus* [2] isolierten Difucolhexaacetat (2,4,6,2',4',6'-Hexaacetoxybiphenyl). Von 3 und 4 konnten keine zur Aufnahme von Spektren ausreichenden Mengen isoliert werden. Aufgrund des DC Vergleiches in verschiedenen Fließmittelsystemen und der Farbreaktion mit Vanillin-H₂SO₄ wird angenommen, daß es sich bei 4 um das Trifucolnonaacetat und bei 5 um das höhere Homologe: Tetrafucoldodekaacetat (A oder B oder beide) handelt, die ebenfalls zuvor aus *Fucus vesiculosus* isoliert worden sind [2].

Anmerkungen—Wir danken der DFG für die Unterstützung dieser Arbeit, dem Cusanus-Werk für ein Promotionsstipendium und der Station Biologique de Roscoff für die Arbeitsmöglichkeit zur Sammlung der Algen.

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XANTHONES AND A CINNAMIC ACID DERIVATIVES FROM *POLYGALA TENUIFOLIA*

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Key Word Index—*Polygala tenuifolia*; Polygalaceae; root; 3,4,5-trimethoxycinnamic acid; xanthones; 1,2,3,7-tetramethoxyxanthone; 1,2,3,6,7-pentamethoxyxanthone; 6-hydroxy-1,2,3,7-tetramethoxyxanthone

Polygalae radix, the root of *Polygala tenuifolia* Willdenow, has been used as an expectorant and sedative similar to *Senega radix*. Early investigations of the plant were focused on the saponins [1], sapogenins [2] and sugars

[3]. We have now isolated 4 fluorescent compounds from an ethereal extract and their structures have been elucidated.

The ethereal extract was separated into soluble and

insoluble fractions in $\text{Ba}(\text{OH})_2$ solution. Column chromatography on Si gel of the substance from the soluble fraction was effected to isolate compounds 1, $\text{C}_{12}\text{H}_{14}\text{O}_5$, mp 129° , and 2, $\text{C}_{17}\text{H}_{16}\text{O}_7$, mp 225° , both of which show blue fluorescent spots on a filter paper under UV light. The insoluble substances were subjected to Al_2O_3 column chromatography to afford compounds 3, $\text{C}_{17}\text{H}_{16}\text{O}_6$, mp 133° and 4, $\text{C}_{18}\text{H}_{18}\text{O}_7$, mp 183° , both of which show white fluorescent spots.

Compound 1 was assigned the structure 3,4,5-trimethoxycinnamic acid, since the UV [λ_{max} 222 nm (ϵ 25 600) and 282 nm (18 000)] and IR [ν_{max} 3100, 3010, 1690, and 1622 cm^{-1}] spectra show the presence of an aromatic ring, and a conjugated carboxylic group. The PMR spectrum [δ 6.76 (s, 2H), 6.36, 7.74 (2H, AB type, $J = 16\text{ Hz}$)] suggests the presence of two equivalent protons on an aromatic ring and protons on a conjugated *trans* double bond. The compound was identified by comparison with 3,4,5-trimethoxycinnamic acid derived from methylation of an authentic sample of sinapic acid.

UV spectra of compound 2, 3, and 4 were quite unlike those of flavonoids, but similar to those of xanthenes [4] which generally show high intensity bands in the region 240–270 nm and relatively low intensity to ca 400 nm. Compound 3, the xanthone isolated from the neutral fraction, gave a yellow colour with conc H_2SO_4 and on adding Mg ribbon with alcoholic HCl. The PMR spectrum shows OMe groups and 4 aromatic protons. One of these aromatic protons appears as a singlet (δ 6.65) and the other 3 are coupled to each other, indicating that one ring of the xanthone is substituted with 3 OMe groups and the other by one. The proton signal (δ 7.60) at lowest field indicates the presence of a C-8H, which is deshielded by the adjacent carbonyl group. The J 's of 0.5 and 1.5 Hz suggests the presence of *meta*, and *para* protons at C-6 and C-5, respectively. On the basis of the spectral evidence and the oxygenation patterns of xanthenes 5 and 6 previously isolated from *Polygala* spp.

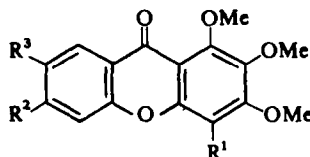
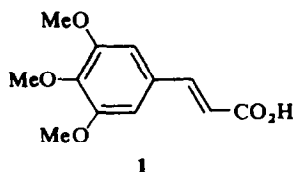
[4–6], compound 3 was assigned the structure 1,2,3,7-tetramethoxyxanthone. This was confirmed by comparison with an authentic sample of the methylation product of 1-hydroxy-2,3,7-trimethoxyxanthone which has previously been reported from the Gentianaceae [7].

One of the isolated xanthenes, 2, was converted into another, 4, by methylation, hence they have the same oxygenation pattern on the skeleton. Their PMR spectra show the presence of 3 aromatic protons as singlets (2: δ 6.67, 6.87, and 7.62. 4: δ 6.68, 6.8 and 7.58), and of 4 and 5 OMe for 2 and 4, respectively, suggesting that both xanthenes are oxygenated at 1, 2, 3, 6, and 7 positions of the skeleton. Comparison of the PMR of both with 3 shows that 1) the signals resonating at the lowest field correspond to C-8H, 2) the signals at δ 6.80–6.87 (C-5H) are reported to be shifted ca 0.4–0.5 ppm upfield by an adjacent OH or OMe group [7, 8] and 3) the signals at δ 6.67–6.68 may be assigned to C-4H. Consequently, the structure of 4 can be represented by 1,2,3,6,7-pentamethoxyxanthone and 2 is the corresponding desmethyl derivative. Further PMR examination in C_6D_6 [4] shows that two signals of the OMe in 2 shift upfield, while 3 signals of OMe in 4 shift upfield, suggesting that a free OH group in 2 can be located at the 3, 6 or 7 position. Furthermore, the UV of 2 after the addition of NaOAc gives a bathochromic shift supporting the presence of the free OH group at the 3 or 6 position [9a, b]. The 1,2,3-triOMe group on the skeleton is suggested for 2 by analogy with congeners. Consequently, the structure of 2 is probably represented by 6-hydroxy-1,2,3,7-tetramethoxyxanthone.

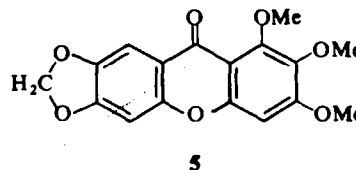
EXPERIMENTAL

IR spectra were recorded in KBr discs; PMR spectra were determined at 100 MHz using TMS as internal standard; UV spectra were taken in MeOH or EtOH; Fluorescence spectra (FS) were measured in MeOH by excitation light at 265 nm. All mps are corrected.

Extraction. Dried and cut roots of *Polygala tenuifolia* Willdenow (10 kg) from China were allowed to stand in Et_2O (54 l) for two weeks at room temp. and then the mixture was warmed for 24 hr at 37° and filtered. The Et_2O extract was evapd to leave an oily residue (1.17 kg), which was separated into a sol. and an insol. fraction in a satd aq. soln of $\text{Ba}(\text{OH})_2$. The sol. part was acidified with 20% HCl and extracted with CHCl_3 , until the



- 2 $\text{R}^1 = \text{H}$, $\text{R}^2 = \text{OH}$, $\text{R}^3 = \text{OMe}$
 3 $\text{R}^1 = \text{R}^2 = \text{H}$, $\text{R}^3 = \text{OMe}$
 4 $\text{R}^1 = \text{H}$, $\text{R}^2 = \text{R}^3 = \text{OMe}$
 6 $\text{R}^1 = \text{R}^3 = \text{OMe}$, $\text{R}^2 = \text{H}$



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organic phase was no longer fluorescent. The combined CHCl_3 extract was concd and chromatographed on a Si gel column Na_2CO_3 soln acidified and re-extracted with CHCl_3 . The CHCl_3 extract was concd and chromatographed on a Si gel column (2.5×40 cm), eluting with CHCl_3 .

Isolation and characterization. The early eluates gave crude crystals (250 mg), which were recrystallized from EtOH to yield 1, as colourless needles, mp 129° ; $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 222, 282, (log ϵ 4.408, 4.255); ν_{max} cm^{-1} : 3100, 3010, 2980, 2950, 2850, 1690, 1622, 1583, 1500, 833; PMR (CDCl_3) δ m: 3.88 (3s, 9H, $3 \times \text{OMe}$), 6.36 (d, 1H, $J = 16$ Hz), 6.76 (s, 2H), 7.74 (d, 1H, $J = 16$ Hz), 9.5 (br, 1H, CO_2H); MS m/e : 238 M^+ , 223, 195, 163; FS nm: 424; (Found C, 60.68; H, 6.07. $\text{C}_{12}\text{H}_{14}\text{O}_5$ requires C, 60.51; H, 5.88%). It was identical with 3,4,5-trimethoxycinnamic acid, which was derived from methylation of sinapic acid (TLC, IR, mmp). The later eluates afforded crude crystals (350 mg), which were recrystallized from EtOH to give 2 as colourless needles, mp 225° ; $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 244, 274, 313, 343 sh (log ϵ 4.62, 4.13, 4.35, 4.10), $\lambda_{\text{max}}^{\text{EtOH} + \text{NaOAc}}$ nm: 240, 272 sh, 320 sh, 360 (log ϵ 4.66, 4.11, 4.07, 4.29); ν_{max} cm^{-1} : 3120, 3010, 2950, 2850, 1640, 1617, 1595, 1475, 1460, 1425, 1415, 1280, 1215, 1205, 1125; PMR (CDCl_3) δ_{ppm} : 3.7 (s, 3H, OMe), 3.8–3.83 (3s, 9H, $3 \times \text{OMe}$), 6.45 (1H, OH), 6.67 (s, 1H, C-4H), 6.87 (s, 1H, C-5H), 7.62 (s, 1H, C-8H); PMR (C_6D_6) δ_{ppm} : 3.07, 3.14, 3.73, 4.1 (each s and 3H, $4 \times \text{OMe}$); MS m/e : 332 M^+ , 317, 302, 289, 274, 259, 167; FS nm: 454; (Found C, 61.27; H, 4.78. $\text{C}_{17}\text{H}_{16}\text{O}_7$ requires C, 61.45; H, 4.85%). The solid which was insol. in $\text{Ba}(\text{OH})_2$ soln was extracted several times with MeOH, until the MeOH extract showed no fluorescence. The combined MeOH extracts were evapd *in vacuo* to give a resinous mass (240 g), which was suspended in 10% Na_2CO_3 soln (12 l) and extracted with Et_2O . The Et_2O soln was concd and chromatographed on a Al_2O_3 column (2.5×40 cm) eluting with Et_2O . The early eluates gave crude crystals (2.1 g), which were recrystallized from EtOH to yield 3, as colourless needles, mp 133° ; $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 242, 256, 280, 310, 355 (log ϵ 4.51, 4.51, 4.45, 4.49, 3.78); ν_{max} cm^{-1} : 3040, 2980, 2950, 2850, 1652, 1615, 1595, 1480, 1470, 1460, 1422, 1360, 1285, 1145, 1030, 815; PMR (CH_2Cl_2) δ_{ppm} : 3.81–3.92 (4s, 12H, $4 \times \text{OMe}$), 6.65 (s, 1H, C-4H), 7.2 (dd, 1H, $J = 1.5, 4.8$ Hz, C-6H), 7.3 (dd, $J = 0.5, 4.8$ Hz, C-5H), 7.6 (dd, 1H, $J = 0.5, 1.5$ Hz, C-8H); MS m/e : 316 M^+ , 301, 273, 258, 151; FS nm: 484; (Found C, 64.52; H, 4.98. $\text{C}_{17}\text{H}_{16}\text{O}_6$ requires C, 64.55; H, 5.10%). It was identical with an authentic sample of

1,2,3,7-tetramethoxyxanthone (TLC, IR, and mmp). The later eluates gave crude crystals, which were recrystallized from EtOH to yield 4, mp 183° ; $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 247, 270 sh, 312, 342 sh (log ϵ 4.43, 3.96, 4.15, 3.86), $\lambda_{\text{max}}^{\text{EtOH} + \text{NaOAc}}$ nm: 248, 272, 312, 343 sh (log ϵ 4.42, 3.95, 4.12, 3.83); ν_{max} cm^{-1} : 3010, 2970, 2950, 2850, 1642, 1620, 1610, 1595, 1510, 1475, 1425, 1280, 1270, 1220, 1130, 1060; PMR (CD_2Cl_2) δ_{ppm} : 3.81–3.85 (5 s, 15H, $5 \times \text{OMe}$), 6.68 (s, 1H, C-4H), 6.8 (s, 1H, C-5H), 7.58 (s, 1H, C-8H); PMR (C_6D_6) δ_{ppm} : 3.33, 3.37, 3.48, 3.79, 4.09 (each s and 3H, $5 \times \text{OMe}$); MS m/e : 346 M^+ , 331, 315, 303, 288, 273, 181; FS nm: 468; (Found: C, 62.47; H, 5.27. $\text{C}_{18}\text{H}_{18}\text{O}_7$ requires C, 62.42; H, 5.44%). It was identical with the Me ether of 2, which was derived from the treatment of 2 with CH_2N_2 -MeOH.

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XANTHONES FROM *LAWSONIA INERMIS*

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Key Word Index—*Lawsonia inermis* (syn. *L. alba*); Lythraceae; lawsone (2-hydroxy-1, 4-naphthaquinone); laxanthone-I (1, 3-dihydroxy-6, 7-dimethoxyxanthone); laxanthone-II (1-hydroxy-3, 6-diacetoxy-7-methoxyxanthone).

Abstract—Two new xanthones isolated from *Lawsonia inermis* have been characterised as 1, 3-dihydroxy-6, 7-dimethoxyxanthone and 1-hydroxy-3, 6-diacetoxy-7-methoxyxanthone and named laxanthone-I and II, respectively.

This communication describes the isolation and structural elucidation of two new xanthones from *Lawsonia*

inermis (Syn. *L. alba*) commonly known as henna. Uses of this plant are described in ref. [1] and previous work